Received: 2015.11.07 Accepted: 2016.10.17 Published: 2017.01.04	Activity of urine arylsulfatase A in brain-dead graft donors is a predictor of early and late graft function					
Authors' Contribution: A Study Design B Data Collection C Statistical Analysis D Data Interpretation E Manuscript Preparation E Literature Search G Funds Collection	Aktywność arylosulfatazy A w moczu dawców narządów ze śmiercią mózgu jako czynnik prognostyczny wczesnej i późnej funkcji przeszczepu Ewa Kwiatkowska ¹ A B C D E F , Leszek Domanski ¹ B D , Joanna Bober ² B D , Krzysztof Safranow ³ C D , Andrzej Pawlik ⁴ D E , Kazimierz Ciechanowski ¹ F G ¹ Department of Nephrology, Transplantology and Internal Medicine, Pomeranian Medical University in Szczecin, Poland ² Department of Medical Chemistry, Pomeranian Medical University, Szczecin, Poland ³ Department of Biochemistry, Pomeranian Medical University, Szczecin, Poland ⁴ Department of Physiology, Pomeranian Medical University, Szczecin, Poland					
Objective:	Summary Human lysosomal arylsulfatase A (ASA) is a member of the sulfatase family. Arylsulfatase A is required to degrade sulfatides. Sulfatides occur in the myelin sheets of the central and peripheral nervous system. In this study we evaluated the urine activity of lysosomal enzyme arylsulfatase A in brain-dead donors as a marker and predictor of short – and long-term renal allograft function.					
Patients/Methods:	We analyzed data from kidney recipients who received organs from brain-dead donors. Data from 40 donors and 68 recipients were analyzed.					
Results:	Results: Urine activity of arylsulfatase A in graft donors correlated positively with creatinine cleara in graft recipients after transplantation: : significantly after 30 days (Rs=0.38, p=0.004) after 3 years (Rs=0.38, p=0.03), and with borderline significance after 14 days (Rs=0.25, p=0 and after one year (Rs=0.23, p=0.07).					
Conclusions:	The results of this study suggest that arylsulfatase A has a protective effect on kidney allograft, and the urine activity of this enzyme in kidney donors correlates positively with graft function.					
Keywords:	arylsulfatase A • graft • kidney					
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Author's address:

Dr Ewa Kwiatkowska, Clinical Department of Nephrology, Transplantology and Internal Medicine, Pomeranian Medical University in Szczecin, Powstańców Wlkp. 72, 70-111 Szczecin, Poland, e-mail: box1a@interia.pl

INTRODUCTION

Donors with brain death (DBDs) are currently the main source of organs in transplantology. The short - and long-term outcomes of allografts obtained from these donors are inferior when compared to living donors [10]. In the case of organs obtained from DBDs delayed graft function more frequently was observed [12]. The explanation for this lies in the process of brain death (BD), itself resulting in a non-physiological environment culminating in significant organ injury prior to organ procurement. The procurement, preservation and reperfusion phases of transplantation result in significant additional injury to the allograft, rendering it susceptible to short – and long-term dysfunction [19]. BD causes complex disturbances of normal homeostatic systems resulting in hemodynamic instability, hormonal impairment, and inflammation [3,15]. BD results in significant cerebral ischemia and intracranial hypertension, resulting in parasympathetic activity followed by severe vasoconstriction [17]. BD is also associated with metabolic changes in cells and tissues.

Human lysosomal arylsulfatase A (ASA) is a member of the sulfatase family. It is synthesized as a 507 amino acid precursor and is processed in the endoplasmic reticulum to yield a 489 amino acid protein [18]. Each sulfatase is characterized by high substrate specificity. The human sulfatases located in the lysosomes are responsible for the degradation of glycosaminoglycans and sulfolipids [14]. Besides their physiological substrates, arylsulfatases also degrade synthetic chromogens and fluorogens [13]. ASA's major natural substrate is cerebroside 3-sulfate, which will accumulate if there is a deficiency in ASA, resulting in a lysosomal storage disorder known as metachromatic leukodystrophy[5].

In this study we evaluated the urine activity of lysosomal enzyme arylsulfatase A in DBDs as a marker and predictor of short – and long-term renal allograft function.

MATERIAL

We analyzed data from kidney recipients who received organs from brain-dead donors in the years 2009-2013. All patients were transplanted in the Department of Transplantology of Pomeranian Medical University and long-term outpatient care took place in the Department of Nephrology, Transplantology and Internal Medicine of Pomeranian Medical University. Data from 40 donors (16 females, 24 males) and 68 recipients (27 females, 41 males) were analyzed. All recipients received triple immunosuppressive therapy: tacrolimus, prednisone, mycophenolate mofetil. We analyzed activity of tubular lysosomal enzyme ASA in urine of brain-dead donors before organ taking. In recipients we examined early and long-term kidney function. Early graft function was assessed as the necessity of hemodialysis treatment in the first week after kidney transplantation. Patients who required hemodialysis in this period were diagnosed with delayed graft function (DGF). Long-term kidney function was assessed as the level of serum creatinine at 1, 2, 3, 4 and 5 years after kidney transplantation. Estimated glomerular filtration rate (eGFR) was determined by the CKD-EPI formula using the calculator of the National Kidney Foundation.

METHODS

Urine samples were collected from all donors for ASA and creatinine analysis before organ procurement. The collected samples were centrifuged at 4000 rpm for 10 min, and urine, without the sediment, was stored at -80° C until the time of analysis. Urine ASA activity was determined using the method described previously by Werner et al. [20]. Urine creatinine concentrations were determined using picric acid as the regent. ASA activity was calculated in relation to creatinine concentration in urine (U/g creatinine).

STATISTICAL ANALYSIS

We used Statistica 10 software (StatSoft, Poland) for statistical analysis. As the Shapiro-Wilk test showed that the distributions of most of the assessed quantitative variables were significantly different from normal (p<0.05), we used non-parametric Spearman's rank correlation coefficient (Rs) for the statistical analysis.

RESULTS

Clinical characteristics of the renal donors are shown in Tables 1, 2.

Urine activity of arylsulfatase A in graft donors correlated positively with creatinine clearance in graft recipients after transplantation: statistically significant after 30 days (Rs=0.38, p=0.004) and after 3 years (Rs=0.38, p=0.03), and with borderline significance after 14 days (Rs=0.25, p=0.08) and after one year (Rs=0.23, p=0.07) (Fig. 1).

DISCUSSION

Brain death triggers a complex cascade of molecular and cellular events including the release of various proinflammatory mediators, leading to a pronounced inflammatory state. The triggering stimulus of this phe-

Ν	Median	Mean±SD	Range			
40	2.76	3.45±2.68	0.13-12.86			
40	48	46.2±11.7	22-75			
40	1.03	1.23±0.82	0.37-5.22			
40	69	80.3±43.6	12-209			
31	70	73.7±13.9	52-120			
	40 40 40	40 2.76 40 48 40 1.03 40 69	40 2.76 3.45±2.68 40 48 46.2±11.7 40 1.03 1.23±0.82 40 69 80.3±43.6			

Table 1. Clinical characteristics of the renal graft donors

N - number of subjects with data available, SD - standard deviation, eGFR - estimated glomerular filtration rate

Table 2. Clinical characteristics of the studied renal transplant recipients

Characteristic	Ν	Median	Mean±SD	Range
Age [years]	68	49	45.3±14.8	18-80
Dialysis before Tx [months]	47	24	27.1±18.2	0-84
Residual diuresis [mL/d]	44	350	611±1000	0-3000
Weight [kg]	46	70.5	69.9±12.9	47-98.5
CIT [hours]	50	21	21.4±9.3	6-42

N - number of subjects with data available, SD - standard deviation, Tx - transplantation, CIT - cold ischemia time

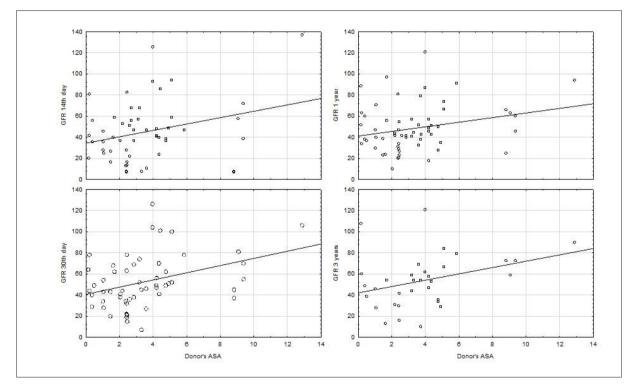


Fig. 1. Correlations between kidney graft donor's urine ASA activity and recipient's eGFR measured 14 days, 30 days, 1 year and 3 years after transplantation

nomenon remains unknown, but it eventually results in endothelial and complement activation, massive cytokine release, hemodynamic impairment and ultimately an immunologically activated organ before transplantation [4]. These changes increase the susceptibility for both ischemia-reperfusion injury and rejection, and may provide an explanation for the inferior results following transplantation of organs from deceased donors as compared with living donors [19]. In our study we analyzed the activity of urine arylsulfatase A in brain-dead graft donors as a predictor of early and late graft function. This activity correlated positively with creatinine clearance after transplantation. These results suggest protective activity of arylsulfatase A in kidney allografts.

Arylsulfatase A is required to degrade sulfatides. Sulfatides, such as galactosylceramide I³-sulfate, occur abundantly in the myelin sheets of the central and peripheral nervous system and in glandular epithelial tissues of mammals. Sulfatides of more complex structure have been found in the kidney [6]. In the human renal cell carcinoma line SMKT-R3, high levels of sulfatides including gangliotriaosylceramide-II³ sulfate were observed [8,9]. In addition, complex sulfatides have been recognized to rank among the strongest ligands for natural killer receptor-p1. This membrane protein, with an extracellular Ca²⁺-dependent lectin domain, is expressed on natural killer cells that display innate immunity [1]. More recently it has been shown that intracellular sulfation of lactosylceramide suppresses the expression of integrins [7] Sulfatides show structural, and possibly physiological similarities to gangliosides. Kidney dysfunction might be correlated with changes in sulfatides, the major acidic glycosphingolipids in this organ. In protein-overload nephropathy mice, the level of sulfatide in serum decreases as the disease progresses. Acute kidney dysfunction lowers the level of sulfatide in serum through downregulation of CST gene expression in lipoprotein-producing organs such as the liver [16]. Reduction of serum sulfatide level in patients with end-stage renal disease was detected prior to induction of hemodialysis therapy [11]. Kidney function itself also seems to be associated with regulation of sulfatide level in serum and lipoprotein-producing organs. Hypoxia in brain death donors may reduce activity of arylsulfatase. Bhattaryya et al. analyzed the effect of hypoxia on arylsulfatase B activity [2]. Hypoxia, like N-acetylgalactosamine-4-sulfatase (arylsulfatase B) silencing, significantly increased the total cellular sulfated glycosaminoglycans and chondroitin-4-sulfate content.

The results of this study suggest that arylsulfatase A has a protective effect on kidney allograft and the urine activity of this enzyme in kidney donors correlates positively with graft function.

REFERENCES

[1] Bezouska K., Yuen C.T., O'Brien J., Childs R.A., Chai W., Lawson A.M., Drbal K., Fiserová A., Pospísil M., Feizi T.: Oligosaccharide ligands for NKR-P1 protein activate NK cells and cytotoxicity. Nature, 1994; 372: 150-157

[2] Bhattacharyya S., Tobacman J.K.: Hypoxia reduces arylsulfatase B activity and silencing arylsulfatase B replicates and mediates the effects of hypoxia. PLoS One, 2012; 7: e33250

[3] Damman J., Seelen M.A., Moers C., Daha M.R., Rahmel A., Leuvenink H.G., Paul A., Pirenne J., Ploeg R.J.: Systemic complement activation in deceased donors is associated with acute rejection after renal transplantation in the recipient. Transplantation, 2011; 92: 163-169

[4] de Vries D.K., Lindeman J.H., Ringers J., Reinders M.E., Rabelink T.J., Schaapherder A.F.: Donor brain death predisposes human kidney grafts to a proinflammatory reaction after transplantation. Am. J. Transplant., 2011; 11: 1064-1070

[5] Halsall D.J., Halligan E.P., Elsey T.S., Cox T.M.: Metachromatic leucodystrophy: a newly identified mutation in arylsulphatase A, D281Y, found as a compound heterozygote with I179L in an adult onset case. Hum. Mutat., 1999; 14: 447

[6] Ishizuka I.: Chemistry and functional distribution of sulfoglycolipids. Prog. Lipid Res., 1997; 36: 245-319

[7] Kabayama K., Ito N., Honke K., Igarashi Y., Inokuchi J.: Suppression of integrin expression and tumorigenicity by sulfation of lactosylceramide in 3LL Lewis lung carcinoma cells. J. Biol. Chem., 2001; 276: 26777-26783

[8] Kobayashi T., Honke K., Kuramitsu Y., Hosokawa M., Miyazaki T., Murata J., Saiki I., Ishizuka I., Makita A.: Cell-surface sulfoglycolipids are involved in the attachment of renal-cancer cells to laminin. Int. J. Cancer, 1994; 56: 281-285

[9] Kobayashi T., Honke K., Kamio K., Sakakibara N., Gasa S., Miyao N., Tsukamoto T., Ishizuka I., Miyazaki T., Makita A.: Sulfolipids and glycolipid sulfotransferase activities in human renal cell carcinoma cells. Br. J. Cancer, 1993; 67: 76-80

[10] Koo D.D., Welsh K.I., McLaren A.J., Roake J.A., Morris P.J., Fuggle S.V.: Cadaver versus living donor kidneys: impact of donor factors on antigen induction before transplantation. Kidney Int., 1999; 56: 1551-1559

[11] Li G., Hu R., Kamijo Y., Nakajima T., Aoyama T., Ehara T., Shigematsu H., Kannagi R., Kyogashima M., Hara A.: Kidney dysfunction induced by protein overload nephropathy reduces serum sulfatide levels in mice. Nephrology, 2009; 14: 658-662

[12] Matas A.J., Humar A., Gillingham K.J., Payne W.D, Gruessner R.W., Kandaswamy R., Dunn D.L., Najarian J.S., Sutherland D.E.: Five preventable causes of kidney graft loss in the 1990s: A single-center analysis. Kidney Int., 2002; 62: 704-714

[13] Németh M., László A., Kovács A., Falkay G.: Lysosomal enzyme activities in frozen, non-cultured chorionic villi for prenatal diagnosis of enzymopathies. Acta Med. Hung., 1992-1993; 49: 143-148

[14] Parenti G., Meroni G., Ballabio A.: The sulfatase gene family. Curr. Opin. Genet. Dev., 1997; 7: 386-391

[15] Schuurs T.A., Gerbens F., van der Hoeven J.A., Ottens P.J., Kooi K.A., Leuvenink H.G., Hofstra R.M., Ploeq R.J.: Distinct transcriptional changes in donor kidneys upon brain death induction in rat: insight in the processes of brain death. Am. J. Transplant., 2004; 4: 1972-1981

[16] Tadano-Aritomi K., Hikita T., Fujimoto H., Suzuki K., Motegi K., Ishizuka I.: Kidney lipids in galactosylceramide synthase-deficient mice. Absence of galactosylsulfatide and compensatory increase in more polar sulfoglycolipids. J. Lipid Res., 2000; 41: 1237-1243

[17] Terasaki P.I., Cecka J.M., Gjertson D.W., Takemoto S.: High survival rates of kidney transplants from spousal and living unrelated donors. N. Engl. J. Med., 1995; 333: 333-336

[18] Von Figura K., Schmidt B., Selmer T., Dierks T.: A novel protein modification generating an aldehyde group in sulfatases: its role in catalysis and disease. Bioessays, 1998; 20: 505-510

[19] Weiss S., Kotsch K., Francuski M., Reutzel-Selke A., Mantouvalou L., Klemz R., Kuecuek O., Jonas S., Wesslau C., Ulrich F., Pascher A., Volk H.D., Tullius S.G., Neuhaus P., Pratschke J.: Brain death activates donor organs and is associated with a worse I/R injury after liver transplantation. Am. J. Transplant., 2007; 7: 1584-1593

[20] Werner M.J., Maruhn D., Atoba M.: Use of gel filtration in the assay of urinary enzymes. J. Chromatogr., 1969; 40: 254-263

The authors have no potential conflicts of interest to declare.