Received: 2012.08.26 Accepted: 2012.10.25 Published: 2012.11.22	Polymorphism of <i>CD36</i> gene, carbohydrate metabolism and plasma CD36 concentration in obese children. A preliminary study*							
 Authors' Contribution: A Study Design B Data Collection C Statistical Analysis D Data Interpretation E Manuscript Preparation F Literature Search G Funds Collection 	Polimorfizm genu <i>CD36</i> a gospodarka węglowodanowa i osoczowe stężenie CD36 u dzieci otyłych. Doniesienie wstępne Monika E. Rać ^{1,0003} , Beata Krupa ²⁰⁰ , Barbara Garanty-Bogacka ²⁰ , Małgorzata Syrenicz ²⁰ Krzysztof Safranow ¹⁰⁰ Violetta Dziedziejko ¹⁰							
	Grzegorz Kurzawski ³ ™, Maria Olszewska ^{1™} , Michał Rać ^{4™} , Dariusz Chlubek ^{1™} ¹ Department of Biochemistry and Medical Chemistry Pomeranian Medical University, Szczecin, Poland ² Independent Laboratory of Propedeutics in Pediatrics Pomeranian Medical University, Szczecin ³ Department of Genetics and Pathomorphology Pomeranian Medical University, Szczecin ⁴ Department of Diagnostic Imaging County Hospital, Szczecin							
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Wprowadzenie:	Receptor CD36 to błonowa glikoproteina uczestnicząca w usuwaniu oxLDL (utlenionych cząstek LDL) z osocza, wiązaniu produktów glikacji białek, patogenezie insulinooporności, cukrzycy typu 2, mikro- i makroangiopatii cukrzycowej. Ekspresja receptora CD36 na makrofagach zwiększa się u pacjentów z hiperglikemią i genetycznie uwarunkowaną otyłością. Jego rola w rozwoju cukrzycy typu 2 i insulinooporności jest niejednoznaczna. Celem pracy było stwierdzenie czy u dzieci z otyłością polimorfizm genu <i>CD36</i> wiąże się z zaburzeniami gospodarki węglowodanowej oraz zmiennością stężenia CD36 w osoczu.									
Materiał/Metody:	Badaniami objęto 60 dzieci w wieku 10–15 lat: 30 z masą ciała >97 centyla i 30 z prawidłową masą ciała. Wykonano pomiary hemoglobiny glikowanej, wysokości, masy ciała, obwodu talii i bioder oraz RR. Obliczono BMI, WHR oraz MAP. Wykonano DTTG z pomiarem stężenia insuliny. Amplikony eksonów 4–6 <i>CD36</i> z przyległymi intronami analizowano metodą dHPLC. Produkty PCR z wykrytymi zmianami były sekwencjonowane. Osoczowe stężenie CD36 ozna- czono z użyciem testu ELISA.									
Wyniki:	Zidentyfikowano dwie intronowe zmiany: IVS3-6 T/C (rs3173798) i IVS4-10 G/A (rs3211892), niesynonimiczną substytucję G367A (Glu123Lys, rs183461468) w eksonie 5 oraz dwie synonimiczne zmiany w eksonie 6: G573A (Pro191Pro, rs5956) i A591T (Thr197Thr, rs141680676). Nie stwierdzono istotnych statystycznie różnic pomiędzy badanymi grupami genotypowymi w żad- nym z morfometrycznych i biochemicznych parametrów.									
Dyskusja:	Brak jest związku polimorfizmów badanego fragmentu <i>CD36</i> z zaburzeniami gospodarki węglo- wodanowej oraz zmiennością stężenia CD36 w osoczu dzieci otyłych. Jednakże ze względu na stosunkowo małą liczebność grup oraz brak danych co do funkcjonalnych efektów badanych po- limorfizmów konieczne są dalsze badania.									
Słowa kluczowe:	CD36 • otyłość • czynniki ryzyka cukrzycy									
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Author's address:	dr n. med. Monika Rac, Pomorski Uniwersytet Medyczny, Katedra Biochemii i Chemii Medycznej, Powstańców Wielkopolskich 72, 70-111 Szczecin, Poland; e-mail: carmon12@gmail.com									

Streszczenie

INTRODUCTION

Diabetes in adults increases the risk of coronary heart disease and cardiovascular complications [1]. Also, insulin resistance in patients with normoglycemia or impaired glucose tolerance is a risk factor for atherosclerosis [9]. Recent studies indicate the involvement of CD36 protein in the pathogenesis of insulin resistance and type 2 diabetes and diabetic micro-and macrovasculopathy [4,5,10,24]. The CD36 receptor is a membrane glycoprotein present on the surface of many cells. It participates in the removal of oxidized LDL (oxLDL) in plasma and the binding protein glycation products [18,20,23]. It was found that glycation of LDL particles more strongly than their oxidation increases the expression of CD36 receptor, oxLDL uptake and accumulation of cholesterol in macrophages. It therefore appears that glycation of LDL particles in diabetes initiates foam cell formation and accelerated atherosclerosis [14]. Some researchers have found that monocytes in subjects with hyperglycemia have increased expression of the CD36 receptor in comparison to those with normal glucose concentration [22]. In addition, patients with mutations in the gene encoding CD36 are more likely to have type 2 diabetes [6]. CD36 receptor expression in macrophages in patients with diabetes is directly proportional to blood glucose level [8]. Increased expression of the CD36 receptor on macrophages was found in genetically determined obesity, which is associated with insulin resistance. This is probably a consequence of disturbances of insulin signaling in these cells [2,17]. On the other hand, in the Japanese population a CD36-receptor defect was not found to be associated with the occurrence of impaired glucose tolerance, insulin resistance, or diabetes [25]. It is possible that in the future plasma circulating soluble CD36 receptor could be added to non-classical cardiovascular risk factors [11]. However, its role in the development of insulin resistance and type 2 diabetes is not clearly understood, and needs further investigations. The available literature contains no reports on the importance of the CD36 receptor in obese children. Thus, for a fuller understanding of the role of CD36 in glucose metabolism pathogenesis, further study is required. The aim of the present study was to search for an association between CD36 gene polymorphism and carbohydrate metabolism disturbances

or the variability of plasma soluble CD36 concentrations in obese children.

MATERIAL AND METHODS

The study included 60 Caucasian children (34 girls and 26 boys) aged 10–15 years: 30 with overweight or obesity (study group) and 30 with normal mass (control group) [12,21]. All patients were treated at the Independent Laboratory of Propedeutics in Pediatrics of Pomeranian Medical University in Szczecin (northwestern Poland) in 2008–2010. The patients were Polish residents. The study complies with the principles outlined in the Declaration of Helsinki and was approved by our institutional Ethics Committee. Informed consent was obtained from each patient. Patients with endocrine or chronic diseases were excluded from the study.

There were no significant differences between the study and control groups as regards age $(11.9\pm3.0 \text{ and } 12.7\pm2.1 \text{ years}, p=0.16; 11.9\pm3.0 \text{ and } 12.7\pm2.1 \text{ years}, respectively, p=0.16) or gender (male 11 and 15, respectively, p=0.20). Each patient's weight, height, waist and hip circumference, and systolic and diastolic blood pressure were measured. The body mass index (BMI), waist-to-hip ratio (WHR), and mean arterial pressure (MAP) were calculated. A fa$ sting blood sample (7 mL) was taken for glycated hemoglobin measurement and DNA extraction. Genomic DNA was isolated as previously described [13]. Moreover, an oral glucose tolerance test (1.75 g glucose/kg body mass, max. dose 75 g) was performed with insulin measurement.

Amplicons of exons 4–6 (region encoding the oxLDL domain) including fragments of introns were studied using denaturing high-performance liquid chromatography (DHPLC) technique as previously described [19]. The PCR products with alterations detected by DHPLC were bidirectionally sequenced using the Applied Biosystems Dye-terminator Cycle Sequencing Ready Reaction kit, according to the manufacturer's protocol. Semi-automated sequence analysis was performed using a 373A DNA fragment analyzer (Applied Biosystems, Foster City, CA). Plasma concentrations of human antigen CD36 (Platelet Membrane Glycoprotein IV) were measured using the commercially available enzyme-linked immunosorbent assay (ELISA) kits (EIAab, Wuhan EIAab Science Co., Ltd., China) according to the manufacturer's protocol.

Differences between subgroups of patients classified according to the intron 3 polymorphism (IVS3-6 T/C) and exon 6 (G573A) were tested with the Mann-Whitney test for quantitative variables and Fisher's exact test for qualitative variables.

RESULTS

Changes detected by DHPLC comprised 2 single nucleotide substitutions in introns (IVS3-6 T/C - rs3173798 and IVS4-10 G/A - rs3211892) and 2 synonymous polymorphisms in exon 6 (G573A - rs5956 and A591T - rs141680676). IVS4-10 G/A alteration was detected in one case in both groups and A591T (Thr197Thr) in two cases in the control group. Moreover, a non-synonymous substitution G367A (Glu123Lys - rs183461468) was detected in case in exon 5. Due to low statistical power of the single polymorphisms (IVS4-10 G/A, A591T, G367A) we analyzed statistically only the associations with IVS3-6 T/C and G573A genotypes. The IVS3-6C allele frequency in the whole children population (10.8%) was similar to that described earlier in the Caucasian populations (6.2% to 11.2%), according to the NCBI dbSNP database. The 573A allele frequency (5.8%) was slightly higher than that described in Caucasians (4.2-4.5%) according to the dbSNP database. Genotype distributions were consistent with the Hardy-Weinberg equilibrium for all sequence changes (p=1).

There were no significant differences between the study and control groups as regards genotype frequency (p=0.74 for IVS3-6 T/C and p=0.34 for G573A). But we identified significant differences between these two groups in CD36 plasma concentration (p=0.02 for IVS3-6 T/C and p=0.05 for G573G). There were no significant differences between the genotype subgroups in terms of any of the clinical, morphometric or biochemical analyzed parameters (Table 1). We found only a tendency (p=0.06) to higher fasting insulin level in IVS3-6 TC heterozygotes than in wild-type homozygotes.

DISCUSSION

No data have been published so far that would suggest an association between variation in the CD36 gene and carbohydrate metabolism disturbances or plasma soluble CD36 concentrations in obese children. Available studies on adults did not analyze changes in the sequence presented in this paper. Leprêtre et al. [16] observed no association between the IVS4-10 G/A alteration and type 2 diabetes in the Caucasian population. There was found a low serum adiponectin level and the associated insulin resistance accompanied by A (-178) C alteration in the promoter of CD36 [16]. In addition there was found an association of low plasma concentrations of adiponectin with a rare nonsense mutation T1079G in exon 10, leading to premature termination of translation and the formation of nonfunctional CD36 receptor protein in patients with type 2 diabetes, including in the Caucasian population [17]. Other authors have often (44%) found in Caucasian adults with type 2 diabetes the C (-3489) T (rs1527479) alteration in the promoter of CD36. The proportion of TT genotype within the study group was 26.5% [3]. By contrast 539AC deletion in exon 6, resulting in a reading frame shift, was indicated as the CD36 gene mutation often associated with the presence of type 2 diabetes in the Japanese population [7].

This report is an attempt to draw attention to the possibility of association of *CD36* gene polymorphism with impaired glucose metabolism and the variability of plasma concentrations of the protein CD36 in obese children, which, in contrast to adults, has not been studied by researchers. It seems reasonable to continue the research and its extension to a larger group of patients to be able to draw better conclusions for the pediatric population.

Our results suggest no association of the analyzed fragment *CD36* polymorphism with impaired glucose metabolism or the variability of plasma concentrations of CD36 protein in obese children. However, due to the relatively small group size and the lack of data regarding the functional effects of polymorphisms studied, further research is needed.

CD36	IVS3-6 T/C (control group)			IVS3-6 T/C (study group)			exon 6 G573A (control group)			exon 6 G573A (study group)		
genotype	TT (n = 23)	TC (n=7)	p- value	TT (n=24)	TC (n=6)	p- value	GG (n=26)	GA (n=4)	p- value	GG (n=27)	GA (n=3)	p- value
Age (years)	10.8±3.3	11.1±2.5	0.67	13.5±2.2	12.8±2.6	0.51	10.8±3.2	11.3±3.0	0.86	13.4±2.2	12.5±2.5	0.56
Gender (% males)	35%	43%	0.68	42%	50%	1.00	35%	50%	0.37	44%	100%	0.22
Weight (kg)	43.6±13.5	43.8±13.0	0.78	76.6±14.2	87.4±25.4	0.33	43.5±13.6	45.0±10.1	0.52	79.5±16.5	72.7±24.9	0.66
Waist (cm)	72.5±10.2	69.1±6.92	0.38	97.1±10.4	108±14.4	0.16	71.5±9.94	71.5±2.29	0.83	100±12.1	93.8±10.0	0.39
Hip (cm)	81.8±10.3	81.7±8.12	0.92	103±8.07	108±15.8	0.53	81.8±10.1	81.3±4.16	0.97	104±10.2	101±10.9	0.57
BMI (kg/m ²)	20.6±2.93	19.5±2.58	0.20	28.7±3.57	33±6.54	0.14	20.3±2.97	20.0±1.71	0.66	29.8±4.74	27.6±2.15	0.61
WHR	0.89±0.08	0.85±0.06	0.31	0.94±0.07	1.00±0.04	0.17	0.88±0.08	0.88±0.07	0.97	0.96±0.07	0.93±0.03	0.22
Systolic BP (mmHg)	110±10	102±9	0.09	122±13	133±15	0.13	108±10	118±4	0.26	125±13	120±20	0.66
Diastolic BP (mmHg)	67±12	58±11	0.67	76±12	87±14	0.11	64±12	75±7	0.22	79±13	67±18	0.22
HR (1/min)	82±14	73±13	0.22	80±12	73±10	0.17	79±15	82±9	0.61	80±12.3	70±3	0.11
MAP (mmHg)	81.6±10.2	72.7±9.90	0.09	91.2±11.9	102±14.4	0.13	78.9±10.6	89.2±5.89	0.18	94.5±12.3	84.4±18.4	0.39
Fasting glucose (mg/dL) OGTT 60 min OGTT 120 min	83.7±6.30 97.7±32.3 96.7±27.0	87.9±0.18 86.8±4.56. 103±0.11.2	0.18 0.78 0.83	85.5±6.91 127±35.1 112±36.0	92.3±18.7 150±5.16 108±11.5	0.89 0.67 0.89	84.0±6.29 97.7±32.3 96.7±27.0	90.5±6.31 86.8±14.2 130±25.0	0.10 0.40 0.50	86.5±10.2 132±32.2 111±31.5	88.3±6.95 150±5.16. 108±11.5	0.52 0.67 0.89
Fasting insulin (µU/mL) insulin 60 min insulin 120 min	8.27±5.17 42.0±32.7 46.9±32.5	13.7±8.72 41.6±27.9. 26.7±25.4	0.06 0.93 0.68	17.2±8.50 162±97.8 187±222	46.2±51.7 194±67.6 89.5±25.3	0.14 0.83 0.67	9.19±4.93 42.0±32.7 46.9±32.5	13.7±15.9 41.6±3.61 26.7±3.42	0.80 1.00 0.82	22.7±25.1 172±87.4 157±188	17.6±4.88 194±67.6 89.5±25.3	0.81 0.83 0.67
HbA1c (%)	5.15±1.31	5.51±0.24	0.70	5.57±0.25	5.51±0.18	0.72	5.21±1.19	5.57±0.19	0.35	5.56±0.25	5.63±0.11	0.48
CD36 (µg/mL)	20.4±12.9	30.0±25.7	0.18	16.8±11.0	11.7±3.8	0.49	23.5±17.9	17.8±6.5	0.80	16.1±10.6	13.0±5.9	0.87

Table 1. Clinical, morphometric and biochemical, parameters of obese and normal-weight children stratified by IVS3-6 T/C and G573A CD36 genotypes

Data are given as mean \pm SD of patients with the indicated genotype. OGTT = oral glucose tolerance test; HbA1c = glycated hemoglobin; CD36 = soluble CD36 protein in plasma; BMI = body mass index; WHR = waist-to-hip ratio; HR = heart rate; MAP = mean arterial pressure.

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The authors have no potential conflicts of interest to declare.